

B3.1 Gas Exchange

Ver. 2

Guiding Questions

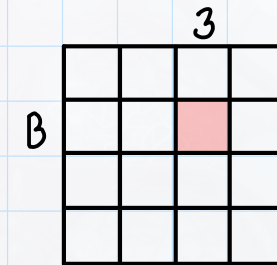
How are multicellular organisms adapted to carry out gas exchange?

What are the similarities and differences in gas exchange between a flowering plant and a mammal?

Linking Questions

How do multicellular organisms solve the problem of access to materials for all their cells?

What is the relationship between gas exchange and metabolic processes in cells?



Theme: Form and Function

Level of Organization: Organisms

Written and drawn by:

PETER MARIER



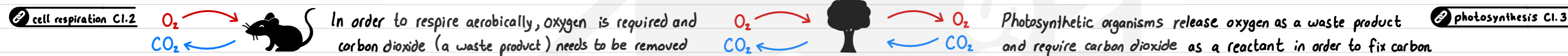
SL Learning Outcomes

B3.1.1	Gas exchange as a vital function in all organisms	Students should appreciate that the challenges become greater as organisms increase in size because surface area-to-volume ratio decreases with increasing size, and the distance from the centre of an organism to its exterior increases.
B3.1.2	Properties of gas-exchange surfaces	Include permeability, thin tissue layer, moisture and large surface area.
B3.1.3	Maintenance of concentration gradients at exchange surfaces in animals	Include dense networks of blood vessels, continuous blood flow, and ventilation with air for lungs and with water for gills.
B3.1.4	Adaptations of mammalian lungs for gas exchange	Limit to the alveolar lungs of a mammal. Adaptations should include the presence of surfactant, a branched network of bronchioles, extensive capillary beds and a high surface area.
B3.1.5	Ventilation of the lungs	Students should understand the role of the diaphragm, intercostal muscles, abdominal muscles and ribs.
B3.1.6	Measurement of lung volumes	Application of skills: Students should make measurements to determine tidal volume, vital capacity, and inspiratory and expiratory reserves.
B3.1.7	Adaptations for gas exchange in leaves	Leaf structure adaptations should include the waxy cuticle, epidermis, air spaces, spongy mesophyll, stomatal guard cells and veins.
B3.1.8	Distribution of tissues in a leaf	Students should be able to draw and label a plan diagram to show the distribution of tissues in a transverse section of a dicotyledonous leaf.
B3.1.9	Transpiration as a consequence of gas exchange in a leaf	Students should be aware of the factors affecting the rate of transpiration.
B3.1.10	Stomatal density	Application of skills: Students should use micrographs or perform leaf casts to determine stomatal density. NOS: Reliability of quantitative data is increased by repeating measurements. In this case, repeated counts of the number of stomata visible in the field of view at high power illustrate the variability of biological material and the need for replicate trials.

HL Learning Outcomes

B3.1.11	Adaptations of foetal and adult haemoglobin for the transport of oxygen	Include cooperative binding of oxygen to haem groups and allosteric binding of carbon dioxide. Students are not required to know myoglobin.
B3.1.12	Bohr shift	Students should understand how an increase in carbon dioxide causes increased dissociation of oxygen and the benefits of this for actively respiring tissues.
B3.1.13	Oxygen dissociation curves as a means of representing the affinity of haemoglobin for oxygen at different oxygen concentrations	Explain the S-shaped form of the curve in terms of cooperative binding. Further biochemical details are given in B3.1.12. An affinity of haemoglobin for oxygen graph is provided in the data booklet.

Gas exchange: the exchange of gases (O_2 and CO_2) between an organism and its surroundings (i.e bringing one in and removing another). ✗ this is a vital function in all organisms

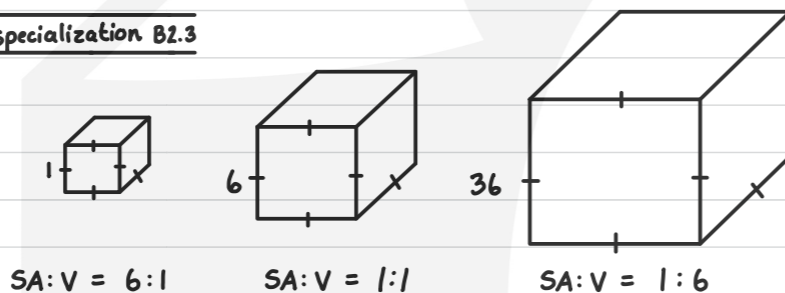


→ process becomes less efficient and more of a challenge the larger an organism becomes due to:

✗ **reduction in surface area : volume ratio**

as an organism increases in size the square-cube law states that its volume (which dictates its metabolic rate and demand) increases faster than its surface area (which dictates its rate of exchange)

cell specialization B2.3



✗ **increased distance for exchange**

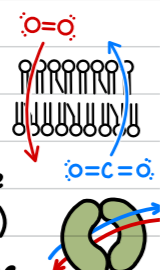
the larger an organism becomes the greater the distance from the center to its exterior, thus making exchange between the innermost cells and the external environment (via diffusion) slow and inefficient



→ in order to maximize the rate of gas exchange, gas-exchange surfaces tend to have the following properties:

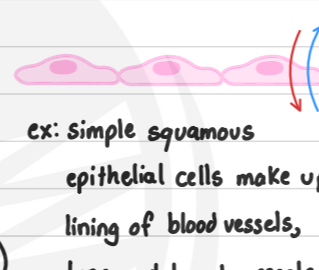
✓ **high permeability**

To quickly and efficiently transport gases across membranes they are freely permeable to O_2 and CO_2 , moving passively via simple diffusion. Vascular plants have pores (stomata) on underside of leaves controlling this exchange



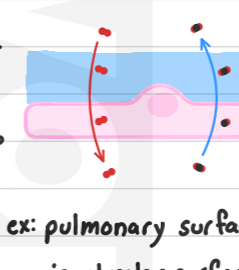
✓ **thin tissue layer**

the shorter the distance gases need to diffuse for exchange, the greater the rate. Thus tissues are adapted to be very thin (1 layer thick and flat in shape)



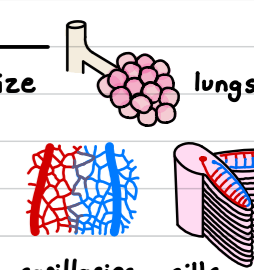
✓ **moist surface**

in order to allow gases to diffuse across plasma membranes, respiratory surfaces in terrestrial animals are covered by a moist film allowing gases to dissolve



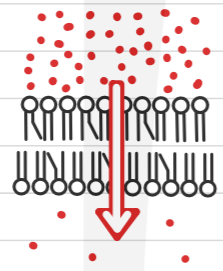
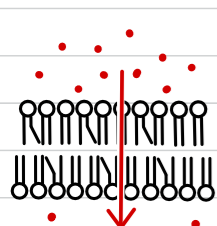
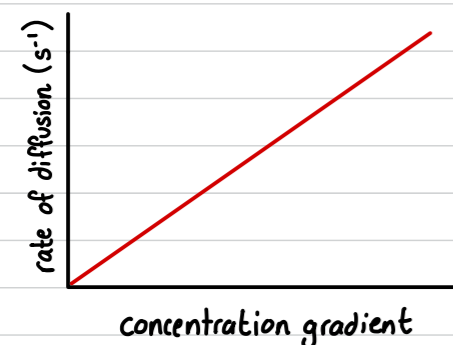
✓ **large surface area : volume**

respiratory surfaces maximize the SA: volume in order to increase the relative area of membrane, where gases will diffuse and be exchanged

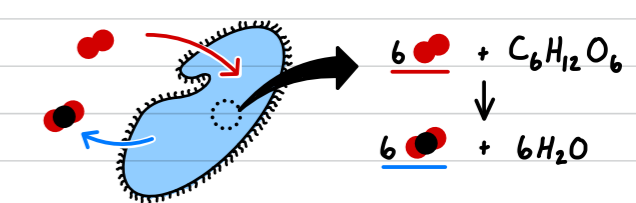


another crucial factor for efficient and continual gas exchange is the maintenance of a large concentration gradient at gas-exchange surfaces

→ concentration gradient is directly proportional to rate of simple diffusion

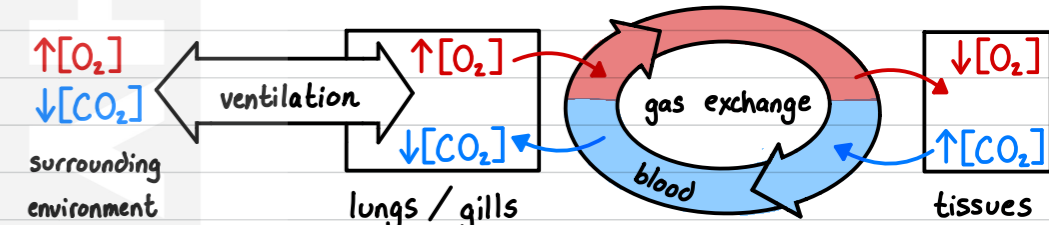


→ in unicellular organisms, such as *Paramecium* a concentration gradient is maintained i.e. high O_2 outside cell, high CO_2 inside cell due to continual aerobic cellular respiration where O_2 is used and CO_2 produced, ensuring passive exchange of both respiratory gases

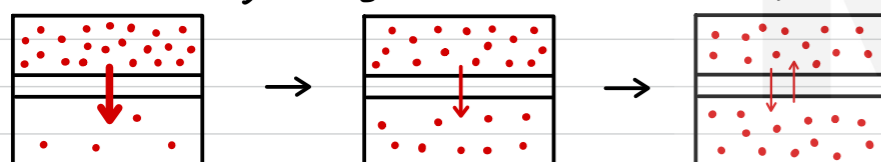


→ in animals, [] gradient is maintained at gas-exchange surfaces:

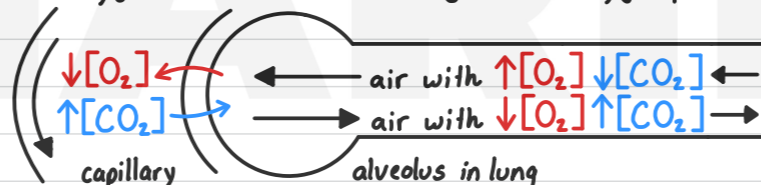
- ✓ ventilating gases in and out of the organism, preventing equilibrium
- ✓ continuous blood flow, preventing equilibrium
- ✓ dense network of blood vessels (capillaries) surrounding tissues: ↑SA



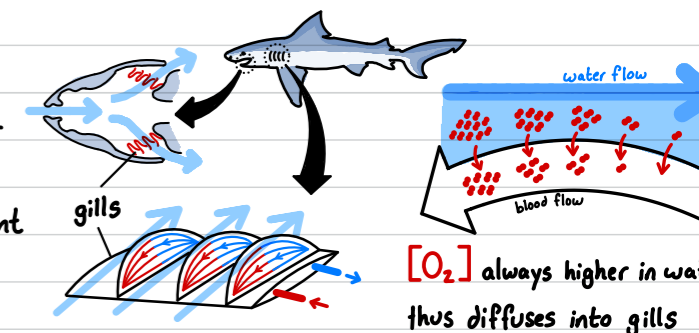
if there is no mechanism to maintain this gradient, diffusion will slow down, eventually resulting in no net movement due to equilibrium



• in mammals, blood flow and ventilation ensures air in lungs is oxygen rich and can be exchanged with oxygen poor blood



• in fish, oxygen rich water is ventilated through the gills. Water flows in the opposite direction of blood in gills, ensuring [] gradient is maintained and that gas exchange occurs continuously



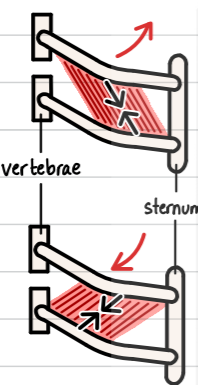
Ventilation: the movement of air in and out of the lungs

↳ In order to do this the volume of air within the chest cavity (thorax) is altered, causing a pressure differential

↳ this is accomplished using **antagonistic muscle pairs**, meaning when one muscle contracts, the other relaxes

Internal and external intercostal muscles

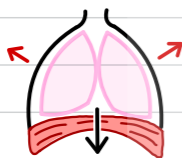
External intercostals contract pulling ribs up and out, internal intercostals relax



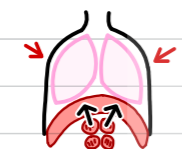
Internal intercostals contract pulling ribs down and in, external intercostals relax

Diaphragm and abdominal muscles

Diaphragm contracts and flattens, pushing relaxed abdominals out

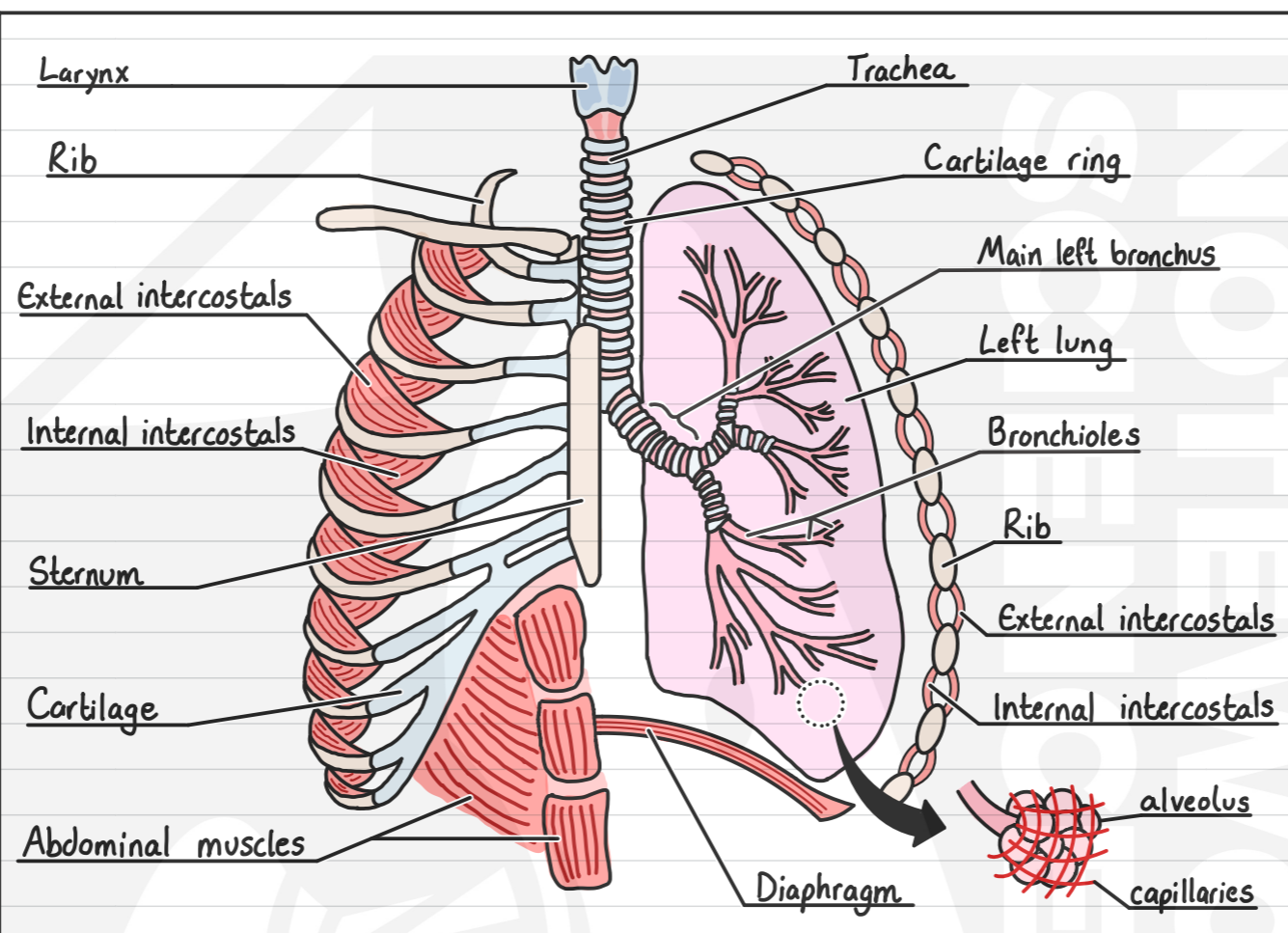


Abdominals contract, pulling ribs down, pushing relaxed diaphragm up (domed)



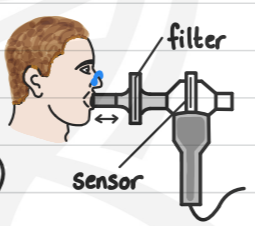
Internal intercostals relax
External intercostals contract
↳ pull ribs up and out
Abdominal muscles relax
Diaphragm contracts
↳ flattens and moves down
↓
Thoracic volume increases
↓
Pressure within thorax decreases below atmospheric pressure
↓
Air flows outside → lungs down pressure gradient

External intercostals relax
Internal intercostals contract
↳ pull ribs down and in
Diaphragm relaxes and domes
Abdominal muscles contract
↳ pushing diaphragm up
↓
Thoracic volume decreases
↓
Pressure within thorax increases above atmospheric pressure
↓
Air flows lungs → outside down pressure gradient

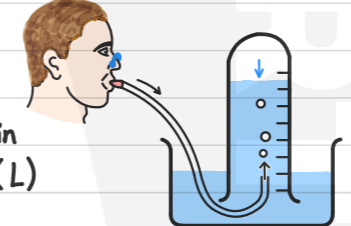


spirometry: test of lung function including lung volumes and ventilation rate

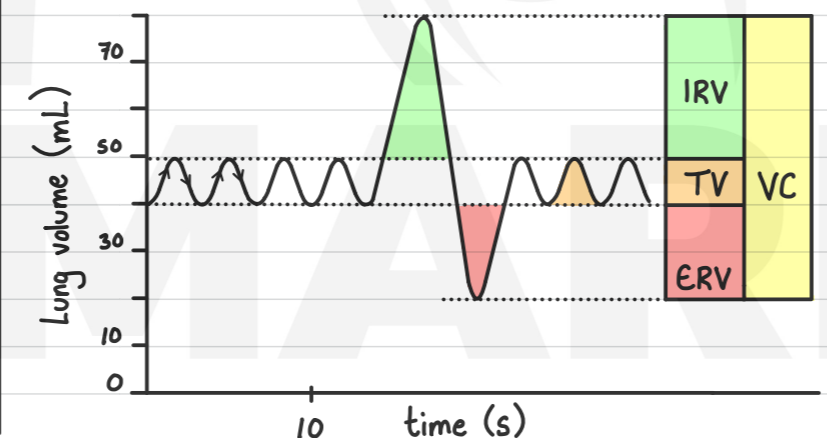
↳ **digital spirometer**
breaths are detected as change in pressure by screens in sensor → flow rate (L/s)



↳ **bell jar spirometer**
air from breaths displaces water within bell jar → volume (L)



Tidal volume (TV): volume of air moved in and out of the lungs in normal ventilation
Inspiratory reserve volume (IRV): maximum of air that can be inhaled after normal inhalation
Expiratory reserve volume (ERV): maximum of air that can be exhaled after normal exhalation
Vital capacity (VC): total amount of air exhaled after a maximum inhalation (IRV + TV + ERV)

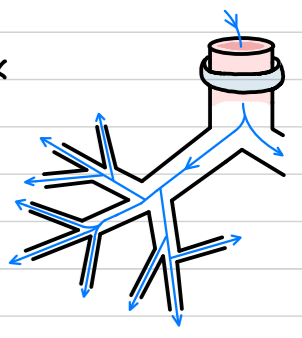


example graph analysis:
TV = 10 mL
IRV = 30 mL
ERV = 20 mL
VC = 10mL + 30mL + 20mL = 60 mL
ventilation rate = 3 breaths / 10s
= 0.3 breaths / s
= 18 breaths / min

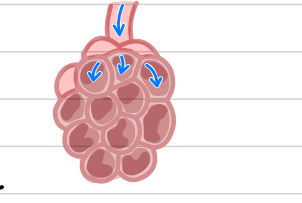
Gas exchange in lungs involves exchanging oxygen from the air in the alveoli with CO₂ from blood in capillaries via simple diffusion

↳ air is exchanged between the outside and the alveolar lung via the airways which has a number of adaptations:

- The airways consist of a branched network of tubes: trachea → bronchi → bronchioles. This allows air to move around lungs evenly. Trachea and bronchi are held open by cartilage rings, resisting collapse. Bronchioles have smooth muscle that adjust airway diameter.



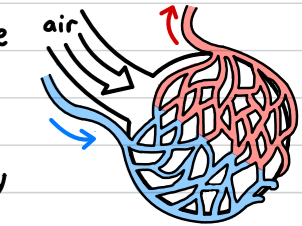
- Each bronchiole terminates in a cluster of spherical alveoli. Despite being small, the large number of these (100s of millions) provide enormous total surface area.



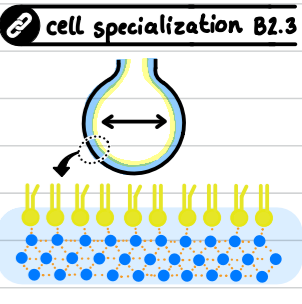
- Each alveolus is surrounded by elastic fibres allowing it stretch during inhalation and recoil during exhalation, forcing air out.



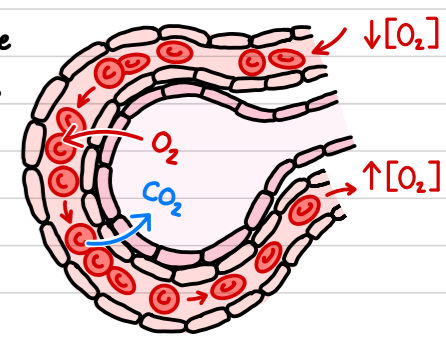
- Each alveolus is surrounded by an extensive capillary bed, providing close access and a very large surface area for gas exchange between air in alveolus and blood in capillary.



- The inside of alveoli are lined with a pulmonary surfactant. As it's moist, it allows O₂ and CO₂ to dissolve and diffuse easier. It also contains phospholipids which decrease surface tension, preventing the alveolus from adhering and collapsing.

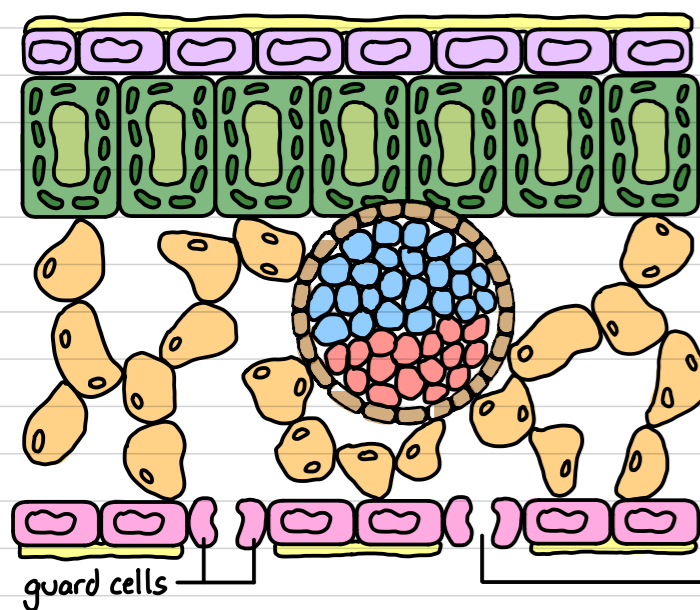


- Both gas exchange tissues in the lung: alveoli and capillaries walls are one cell thick, providing a very small diffusion distance from the inside of the lung to blood, greatly increasing the rate of gas exchange between them.



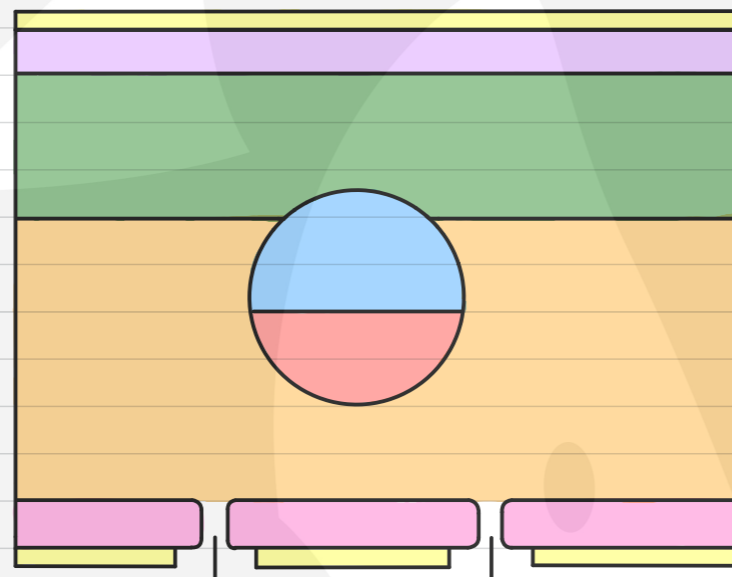
In vascular plants (angiosperms, gymnosperms, ferns) gas exchange occurs in the leaves

transverse cross-section of a dicotyledonous leaf:



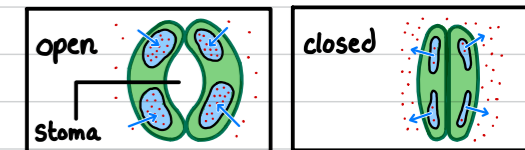
- waxy cuticle
- upper epidermis
- palisade mesophyll
- vein
 - xylem
 - phloem
- spongy mesophyll
- lower epidermis
- stomata
- guard cells

plan diagram of transverse of a dicotyledonous leaf:



leaf tissue gas exchange adaptations:

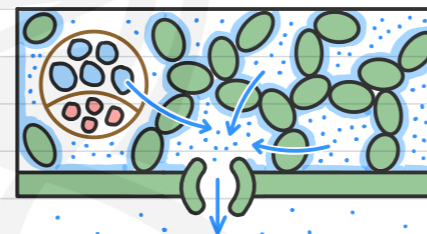
- **waxy cuticle**: the top of leaves as well as the underside (except for stomata) is coated by a layer of lipid wax which is hydrophobic and has low permeability to gases. This ensures that water loss and gas exchange is occurring at the stomata (which can be opened or closed)
- **epidermis**: secrete wax which forms waxy cuticle. Acts as protective layer. Lower epidermis has guard cells which form stomata and regulate gas exchange and water loss
- **spongy mesophyll**: contains many air spaces which provides a large moist surface area for diffusion and gas exchange (CO₂ in, O₂ out for photosynthesizing palisade mesophyll tissue)
- **veins (vascular bundle)**: water is carried in Xylem vessels to leaves for use in photosynthesis and primarily to replace those lost via transpiration. Phloem transports nutrients such as sugars
- **Stomatal guard cells**: pair of cells which regulate gas exchange and water loss via diffusion by either becoming turgid and forming stoma or flaccid and closing



Transpiration - loss of water vapour from leaves of a plant by evaporation from mesophyll air spaces, diffusing out via open stomata

↳ process occurs as a consequence of gas exchange in the leaf, i.e. so long as stomata are open and exchanging O₂ for CO₂:

- ① water delivered to leaf via xylem, keeping spongy mesophyll moist for gas exchange
- ② water within spongy mesophyll air spaces evaporates into water vapour
- ③ increases the relative humidity in air spaces until air is saturated
- ④ water diffuses out of leaf via stomata, dropping humidity within air spaces



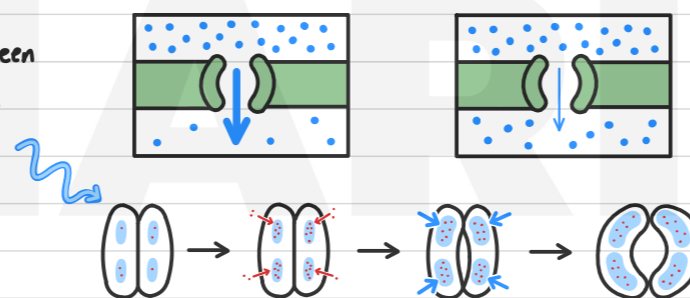
↳ rate of transpiration (i.e. water loss) is impacted by several environmental factors:

● **temperature**: **positive relationship**: higher temperatures mean more kinetic energy to break hydrogen bonds between water molecules ∴ more evaporation and loss via diffusion. * At extreme temperatures, stomata may close to prevent excessive loss

● **wind speed**: **positive relationship**: higher wind speeds displaces more water vapour surrounding leaf away, reducing its relative humidity ∴ greater [] gradient between water vapour inside and outside the leaf ∴ greater rate of diffusion

● **humidity**: **negative relationship**: higher humidity decreases [] gradient between water vapour inside and outside the leaf ∴ lower rate of diffusion

● **light intensity**: **positive relationship**: guard cells swell in response to blue light, opening their stoma to allow gas exchange for photosynthesis * crucial factor as transpiration ceases in absence of light

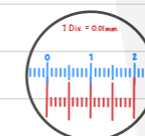


stomatal density: number of stomata per unit area (mm² or μm²) of a leaf surface.

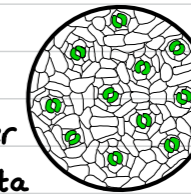
Procedure: use micrograph or take a cast of the lower epidermis of a leaf

- ① spread clear nail polish on section
- ② once dry, place clear tape overtop and press down
- ③ Pull tape off leaf and stick on glass slide

Analysis: view leaf lower epidermis micrograph or casting under microscope at high power



determine the actual field of view diameter using stage micrometer and then count stomata



ex: at 400x, F.O.V. diameter = 0.56mm
 F.O.V. area = π(0.28mm)² = 0.25mm²
 number of visible stomata / area = 11 / 0.25mm² = 44 stomata per mm²

NOS: mean stomatal density should be calculated from multiple micrographs or casts to increase reliability and to account for natural biological variation among specimens

potometer

Diagram of a potometer setup. A plant stem is inserted into a reservoir of water. The stem is connected to a tube that leads to a ruler. An air bubble is trapped in the tube. As the plant transpires, it draws water from the reservoir, which pulls the air bubble along the ruler. The distance the bubble moves over time is measured to determine the transpiration rate.

transpiration draws water, pulling bubble

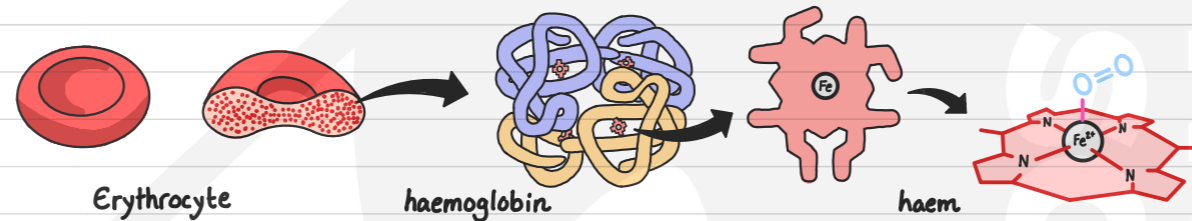
distance bubble moved in 1 min = 0.51mm

transpiration rate = $\frac{\text{volume water (mm}^3\text{)}}{\text{time (min)}}$

determine distance air bubble moves over time

$= \frac{\pi r^2 d}{t} = \frac{\pi (1.5\text{mm})^2 (0.51\text{mm})}{1\text{min}} = 3.6 \frac{\text{mm}^3}{\text{min}}$

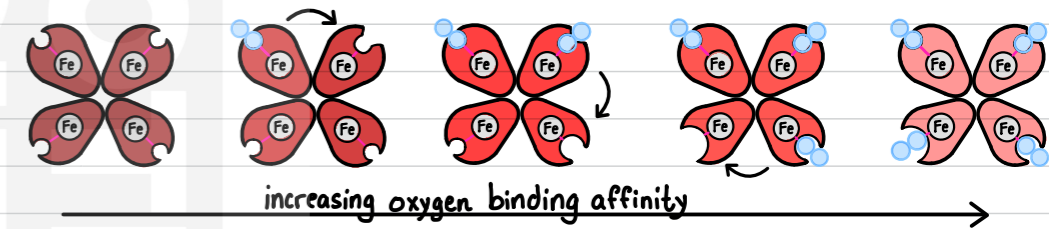
Erythrocytes (red blood cells) are adapted to carry respiratory gases (mainly O_2) through the blood stream by being packed full of the protein haemoglobin. *cell structure A2.2*



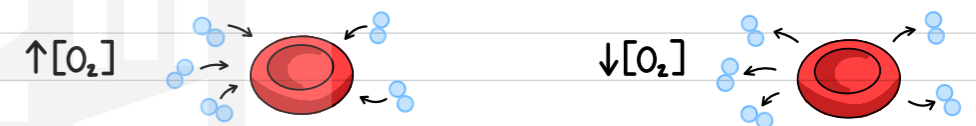
Haemoglobin is a conjugated quaternary protein composed of four subunits - each comprising a polypeptide and a haem group with a bound iron atom which binds reversibly with oxygen. *proteins B1.1*

Cooperative binding of haemoglobin

oxygen (O_2) binding at each of the four haem sites does not occur simultaneously but cooperatively: binding the first oxygen is difficult, but once an oxygen binds it induces a conformational change to the neighbouring chain within the haemoglobin, facilitating subsequent oxygen binding. \therefore binding the first, second, third, and fourth oxygen molecule to haemoglobin becomes progressively easier and conversely, dissociation of oxygen reduces affinity in other haem groups making unbinding progressively easier. \therefore 2 most probable states are Hb with no oxygen and with four

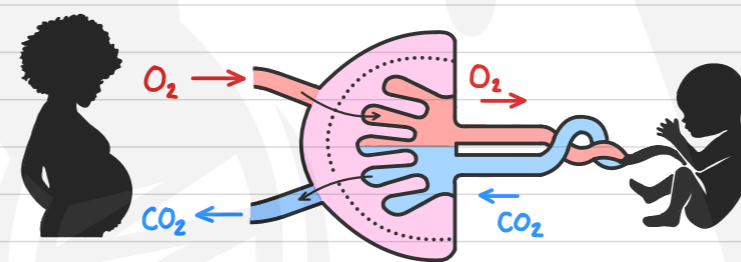


this is adaptive: haemoglobin has a higher affinity for O_2 in oxygen-rich areas (such as the lung) \rightarrow promoting oxygen loading. haemoglobin has a lower affinity for O_2 in oxygen-poor areas (such as muscles) \rightarrow promoting oxygen unloading.



Adaptations of foetal haemoglobin

reproduction D3.1



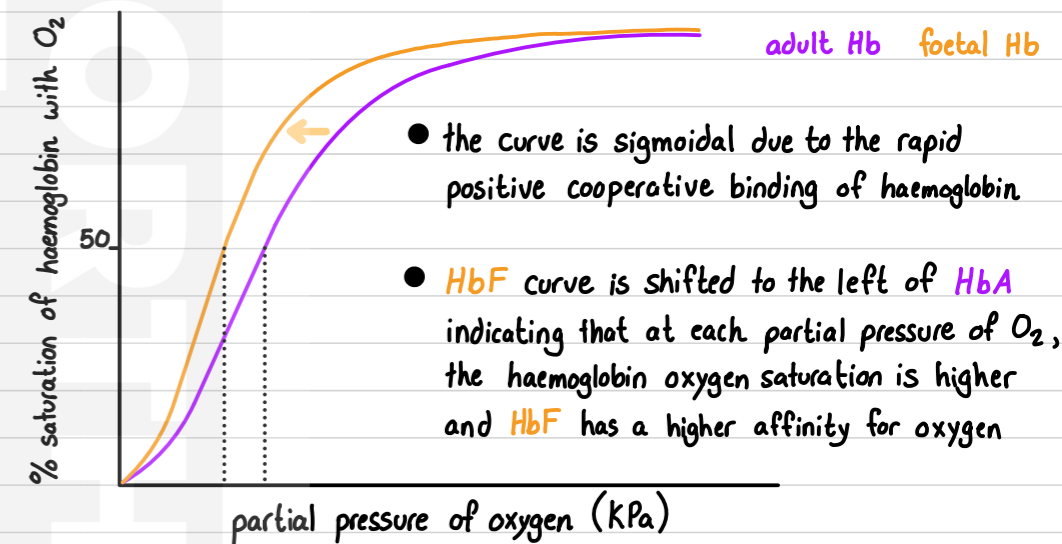
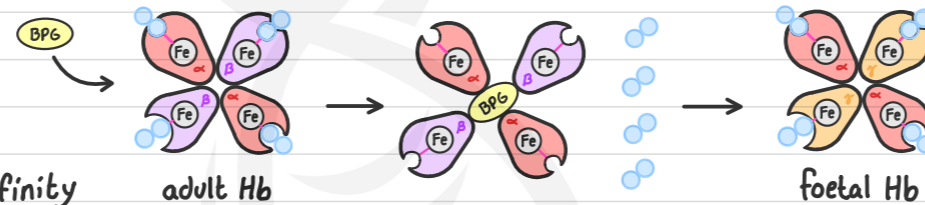
Oxygen dissociation curve: plots the oxygen saturation of haemoglobin over different partial pressures of oxygen. ** found in data booklet*

During pregnancy, the foetus receives O_2 from the mother via the placenta where the mother's red blood cells unload O_2 in the placenta where it diffuses and binds to foetal red blood cells and is delivered via umbilical cord

Problem: how/why does the oxygen bound to the mother's haemoglobin unbind, diffuse and then bind to foetal haemoglobin?

Solution: foetus uses a different haemoglobin (foetal haemoglobin, HbF) which has a higher oxygen affinity than adult haemoglobin

a molecule (2,3-BPG) produced in erythrocytes binds to adult Hb, lowering its O_2 affinity and promoting unloading. HbF has 2 gamma subunits instead of 2 beta subunits which bind 2,3-BPG poorly, allowing it to have higher O_2 affinity

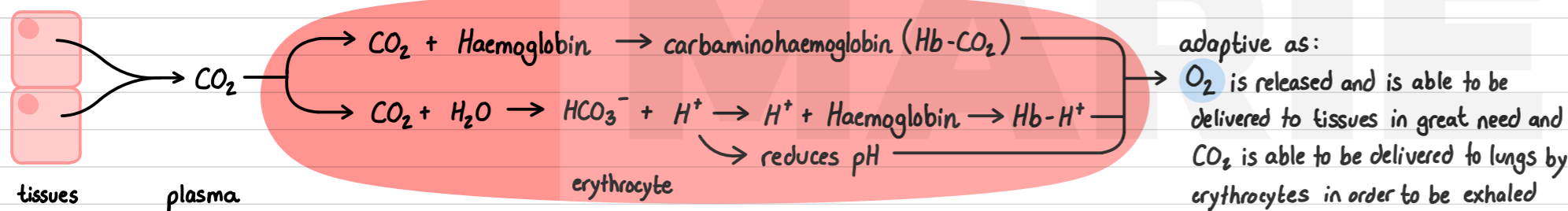


- the curve is sigmoidal due to the rapid positive cooperative binding of haemoglobin.
- HbF curve is shifted to the left of HbA indicating that at each partial pressure of O_2 , the haemoglobin oxygen saturation is higher and HbF has a higher affinity for oxygen

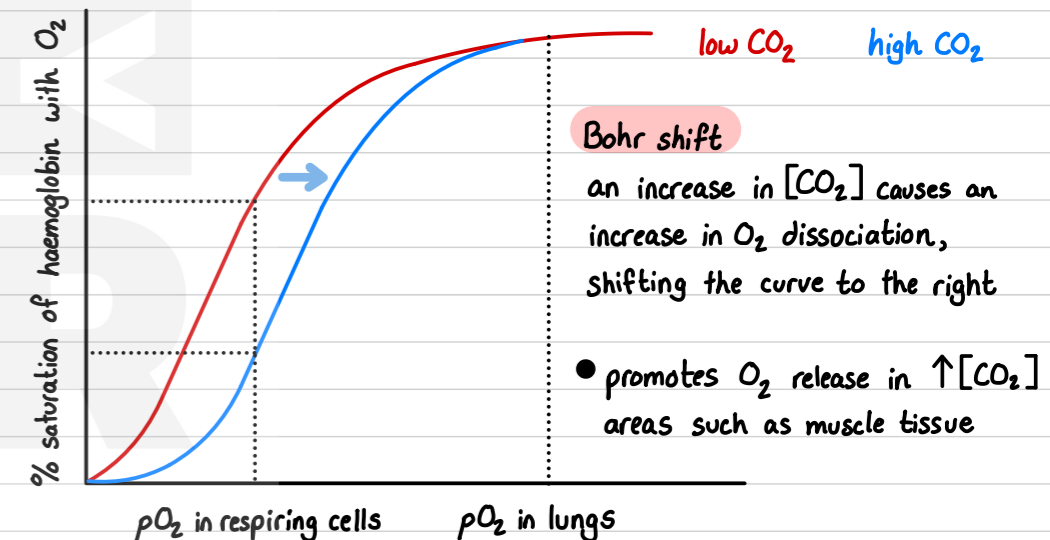
Impact of carbon dioxide - Bohr shift

aerobically respiring tissues produce CO_2 which is released into the blood \rightarrow decreases haemoglobin's affinity for oxygen \rightarrow promotes oxygen offloading

Other substances can bind onto haemoglobin at a site other than the haem (allosteric binding) which causes a change in its conformation



adaptive as: O_2 is released and is able to be delivered to tissues in great need and CO_2 is able to be delivered to lungs by erythrocytes in order to be exhaled



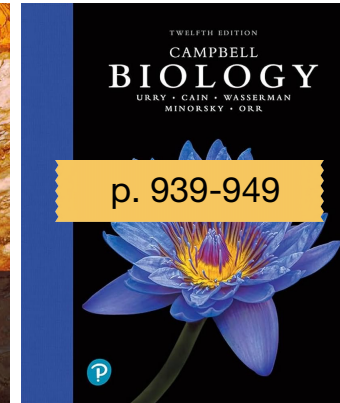
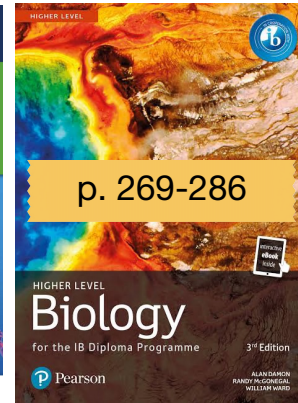
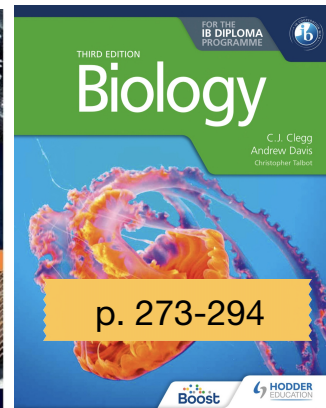
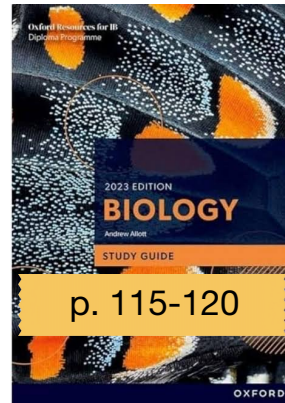
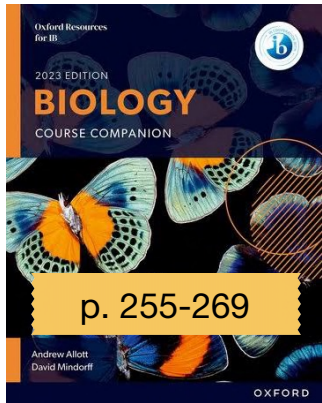
- Bohr shift**
an increase in $[CO_2]$ causes an increase in O_2 dissociation, shifting the curve to the right
- promotes O_2 release in $\uparrow [CO_2]$ areas such as muscle tissue

Resource Links

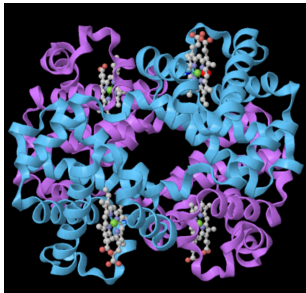
each resource is hyperlinked



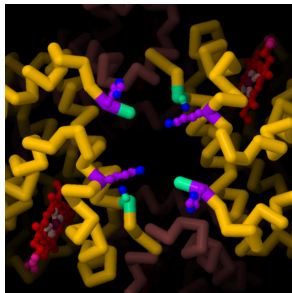
Textbooks



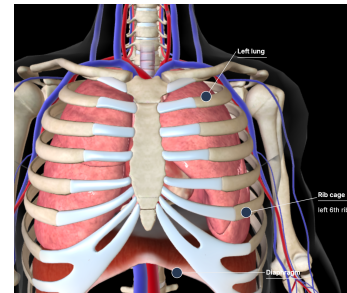
3D models



Adult Haemoglobin



Fetal Haemoglobin



Ventilation

Articles

Demirci, S., Leonard, A., Essawi, K., & Tisdale, J. F. (2021). CRISPR-Cas9 to induce fetal hemoglobin for the treatment of sickle cell disease. *Molecular therapy. Methods & clinical development*, 23, 276–285. <https://doi.org/10.1016/j.omtm.2021.09.010>

Rao, A. A., & Johncy, S. (2022). Tennis courts in the Human Body: A review of the misleading metaphor in medical literature. *Cureus*. <https://doi.org/10.7759/cureus.21474>

Simulators / Interactives

